

# Hydrotropism: analysis of the root response to a moisture gradient

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Running title: Hydrotropic root response to a moisture gradient

## Summary

Hydrotropism is a genuine response of roots to a moisture gradient to avoid drought. An experimental system for the induction of hydrotropic root response in Petri dishes was designed by pioneering groups in the field. This system uses split agar plates containing an osmolyte only in a region of the plate in order to generate a water potential gradient. *Arabidopsis* seedlings are placed on the MS agar plate so that their root tips are near the junction between plain MS medium and the region supplemented with the osmolyte. It elicits a hydrotropic response in *Arabidopsis* roots that can be measured as the root curvature angle.

**Keywords:** hydrotropism, water potential gradient, root curvature angle, moisture gradient, root growth, sorbitol, ABA, *Arabidopsis*

## 1. Introduction

Drought is a major environmental stress that affects plant growth and has a serious impact on agriculture. Plants have developed a range of mechanisms to overcome drought stress including stomatal closure, regulation of gene expression, accumulation of osmoprotectants or compatible solutes and modulation of growth (1). Hydrotropism forms part of the drought-avoidance plant response and is a mechanism by which roots change growth direction according to differences in the water potential of the soil. Elucidating the molecular mechanism of hydrotropism in roots is important both for understanding plant adaptation to soil moisture gradients and improving crop productivity. Root growth is strongly directed by gravity and also by other tropisms like photo-, thigmo- or hydrotropism (2). The study of this latter process is complicated by the fact that gravitropism and hydrotropism influence each other and make it difficult to observe hydrotropism in isolation. However, since roots of the pea mutant *ageotropum* are agravitropic but hydrotropic both sensing pathways must operate through independent mechanisms (3). Additionally, the non-hydrotropic *Arabidopsis* mutants *no hydrotropic response (nhr1)* and *mizu-kussei (miz1)* show a normal root

gravitropic response (4, 5). Unlike gravitropism, where the central role of auxin in its regulation has been well described, the molecular mechanisms that regulate hydrotropism are still largely unknown. Screening procedures in model plant species are needed for genetic dissection of complex biological processes. Therefore, a protocol that allows the analysis of the hydrotropic response in *Arabidopsis* seedlings can help to isolate key genes involved in this response. Takahashi *et al.* (6) and Eapen *et al.* (4) described two methods to measure the hydrotropic response in *Arabidopsis* seedlings. Both methods are based on the same principle of challenging roots with a water potential gradient that will ultimately trigger root bending. Here we describe one of them based on the use of split agar plates that contain two media with different water potential. The use of these strategies has allowed the isolation of mutants that lack a normal hydrotropic response such as *miz1*, *miz2/gnom*, *ahr1* and *nhr1* (6-8, 4), or the identification of mutants that show an enhanced hydrotropic response, such as the *pp2c* quadruple mutant impaired in the protein phosphatases type 2C that negative regulate ABA signalling (9). As a result, it has been identified a gene, MIZ1, that plays an essential role for hydrotropism and is expressed in the columella cells of the root cap (6). It has been also found that GNOM-mediated vesicular trafficking and ABA signalling through PYR/PYL/RCAR receptors is required for hydrotropism in *Arabidopsis* (7, 9). Finally, in addition to ABA (5, 9), cytokinins also play a role in root hydrotropism (8).

## **2. Materials:**

- D-Sorbitol
- Murashige and Skoog (MS) basal salt macronutrient solution (10)
- 120 x 120 x 17 mm square polystyrene Petri dishes
- 3M Micropore surgical tape
- Growth chamber in darkness
- Imaging system (optional, if time course analysis is intended): IR camera system (11)
- NIH image software ImageJ v1.37

### 3. Methods:

#### 3.1. Growth of *Arabidopsis* seedlings

- 1- Sterilise *Arabidopsis* seeds by treatment with 50% sodium hypochlorite for 5 minutes followed by four washes with sterile distilled water and stratify them in the dark at 4°C for 3 days.
- 2- Sow the seeds on plates containing half-strength (0.5x) MS medium, pH 5.6-5.8 and solidified with 1% agar (0.5xMS plates).
- 3- Seal the plates with 3M Micropore surgical tape and put them vertically in a growth chamber with 24h light at 22°C for 5-6 days.

#### 3.2. Preparation of plates that establish a water potential gradient

- 1- The osmotic pressure ( $\pi$ ) of an ideal solution can be approximated using the Morse equation  $\pi = iMRT$ , where  $M$  is the molarity of the solute. Sorbitol is a common solute in laboratory that lacks secondary effects and we use it to generate a water potential gradient in split agar plates. For instance, adding 0.4 M sorbitol as solute lowers the water potential by -1 MPa. Split agar plates are generated by filling the upper region of the square Petri dish with medium containing 0.5xMS salts and the lower region with 0.5xMS salts + 0.4 M sorbitol (**Fig. 1**). Thus, a water potential gradient will be generated between the two different media. Root tips will be located at the border between both media and the root curvature will be measured within 12 h (**Fig. 1**). The water potential gradient between both media becomes smaller over time, but exists for a substantial portion of the experimental period.
- 2- The split agar plates must be prepared just before the experiment. We use a template placed below the plate in order to guide the diagonal cutting of the medium with a scalpel (**Fig. 1**). Next, we remove the 0.5xMS medium from the lower part of

the dish and we replenish the empty part of the plate with 0.5xMS medium supplemented with 0.4 M sorbitol.

- 3- Use the same template to guide the transfer of *Arabidopsis* seedlings grown on vertical plates to the plates with a water potential gradient so that their root tips are 2-3 millimetres above the border between the two media (blue line in **Fig. 1**). Seedlings should be transferred only to the 3 central rectangles. According to the size of the square Petri dishes, 5-6 seedlings of each genotype can be transferred to one of the rectangles in such a way that each plate contains up to 3 different genotypes.
- 4- At least three plates per experiment are needed. In order to avoid a positional effect, the order of the genotypes on the plates should be varied. The two rectangles close to the edges of the plate should be left empty as diffusion of water and sorbitol could be affected by boundary effects.
- 5- Seal the plates with 3M Micropore surgical tape to allow gas exchange.

### 3.3. Root curvature measurement

- 1- Place the plates vertically in a growth chamber at 22°C in darkness and take images of the plants after 8-13 h. Performing the experiment in darkness eliminates the effect of phototropism on root angle, although it has been described that the hydrotropic curvature of light-grown seedlings is notably higher than in dark-grown seedlings (12). In wild type plants root curvature generated in response to the moisture gradient can be observed from the first hour onwards; however, after 8-13 h the angle values of root curvature will be higher, which will make it easier to distinguish between altered and normal hydrotropic response (**Fig. 2A, B**). If detailed analysis of the development of the hydrotropic root curvature over time is intended, images should be taken with an automated IR camera system.
- 2- Use Image J to measure the curvature angle of the primary root (**Fig. 2A**).

#### 4. Notes:

- 1- After the preparation of the plates, the water potential gradient will decrease over time (6). For this reason, plates must be prepared just before the start of the experiment and angle measurements should be taken after 8-13 h.
- 2- While setting up the experiment, maintain plants in vertical orientation as much as possible. As roots tend to grow in the direction of gravity, keeping them in the same initial position will avoid unnecessary perturbations in root growth.
- 3- When preparing the plates take special care of working on a level surface to ensure that the level between the two media is similar. We use a spirit level to ensure the flow laminar hood where media are poured is really on the same level. Additionally take care to obtain a horizontal surface in the border between the two media that generate the water potential gradient. The transition between both media should be as flat as possible since irregular surfaces between both media could alter the root hydrotropic response.
- 4- It is necessary to include in the experiment control 0.5xMS plates lacking the water potential gradient. As shown in **Fig. 2C**, seedlings growing on control plates follow the gravity vector while those growing on plates that contain a water potential gradient bend and avoid the low water potential area. Once the measurement of root curvature is done, seedlings can be left growing for 2-3 days to observe if the response persists in time (**Fig. 2D**).
- 5- As temperature affects osmolyte diffusion and in order to keep the water potential gradient as steep as possible, cool down the 0.5xMS medium supplemented with sorbitol as much as possible before pouring.
- 6- Select seedlings of the same developmental stage, which should have similar root growth rates. Under our growth conditions (22°C in darkness) Col-0 plants show a root curvature of 30-40 degrees after 12 h. In addition to the wild type control, the use

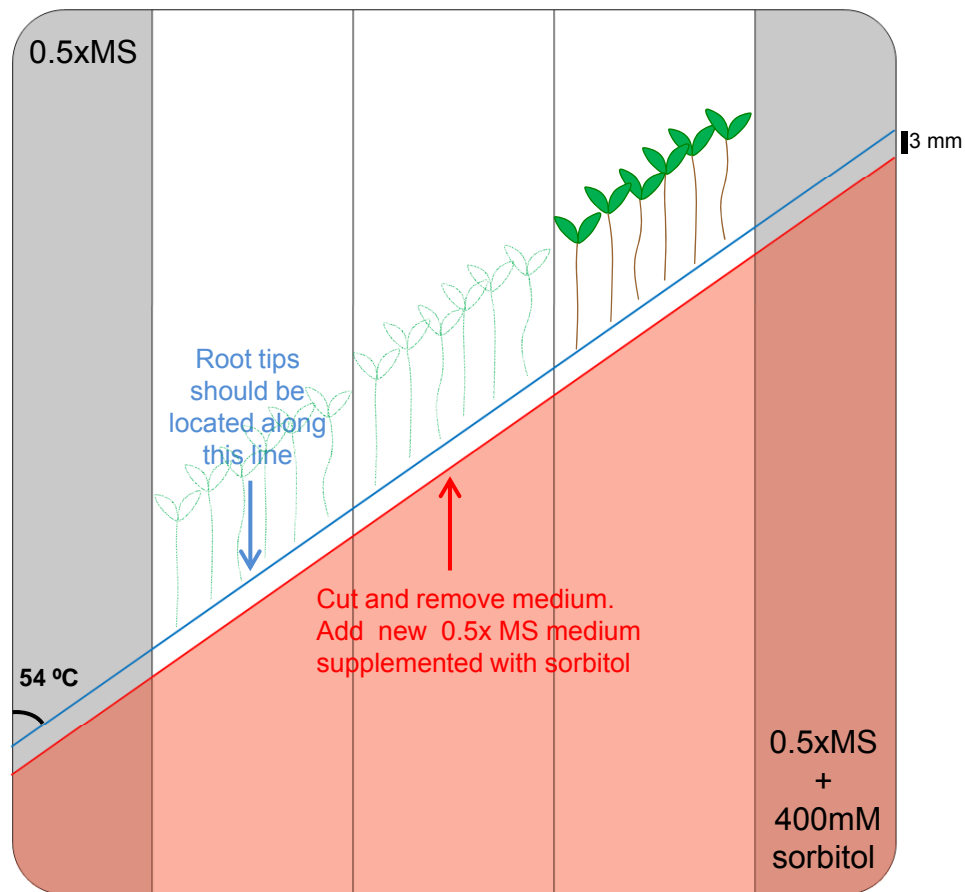
of mutants that lack or show enhanced hydrotropic response is strongly advisable (Fig. 2D).

## Acknowledgment

Funding was provided by a Marie Curie Intra-European Fellowship to R.A and grants from the Ministerio de Ciencia e Innovacion, Fondo Europeo de Desarrollo Regional and Consejo Superior de Investigaciones Cientificas to P.L.R. (BIO2011-23446) and the Biotechnology and Biological Sciences Research Council to D.D and M.J.B.

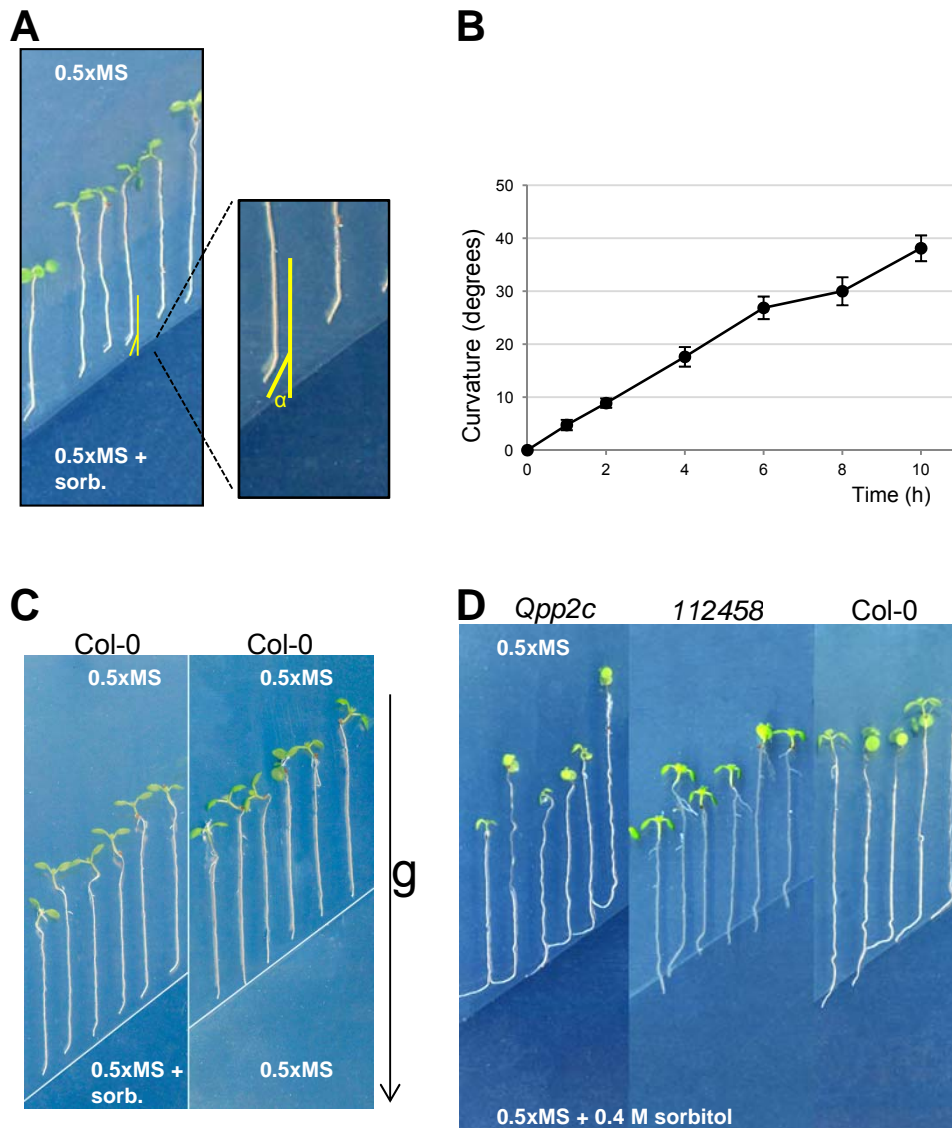
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**Figure 1.** Schematic representation of the split agar plate used for the analysis of the root hydrotropic response. The upper part of the plate contains 0.5xMS medium while the lower part contains 0.5xMS medium supplemented with 0.4 M sorbitol, which generates a water potential gradient. The plate is vertically divided in 5 rectangles. Seedlings should be located in the 3 central rectangles.





**Figure 2.** Root hydrotropic response of 6-day-old *Arabidopsis* seedlings in plates where a water potential gradient has been established. **(A)** Measurement of the root curvature angle ( $\alpha$ ) after 10 h of transferring the seedlings to split agar plates to estimate the hydrotropic response. **(B)** Time course of the hydrotropic response in Col-0 *Arabidopsis* seedlings. **(C)** Root bending only occurs in Col-0 seedlings after transferring them to plates with a water potential gradient. Otherwise, they follow the gravity (g) vector. **(D)** Root hydrotropic response of the ABA-hypersensitive mutant impaired in four clade A PP2Cs (*abi1-2 hab1-1 pp2ca-1 abi2-2*, abbreviated as *Qpp2c*) and the ABA-insensitive mutant impaired in six PYR/PYL ABA receptors (*pyr1 pyl1 pyl2 pyl4 pyl5 pyl8*, abbreviated as *112458*). 6-day-old seedlings were transferred to plates with a moisture gradient and photographs were taken after 3 days. An enhanced hydrotropic response was observed in *Qpp2c*, whereas a lack of response was found in *112458* compared to Col-0 wt seedlings.